

## Magnesium inhibition of skeletal muscle ryanodine receptors modified by DIDS, ryanodine and ATP

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In skeletal muscle the activity of ryanodine receptor (RyR) calcium release channels in the sarcoplasmic reticulum is regulated by the dihydropyridine receptor (DHPR) voltage sensors in the t-tubule membrane.  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  and ATP are potent intracellular regulators of RyRs. The effects of these substances on isolated RyRs are well characterised yet it is not clear how they regulate RyR opening under voltage-sensor control. RyRs are activated by  $\mu\text{M}$  cytoplasmic  $\text{Ca}^{2+}$  and mM ATP while physiological  $[\text{Mg}^{2+}]$  (~1 mM) in the cytoplasm fully inhibits them. It is proposed that during muscle contraction, DHPRs transiently relieve  $\text{Mg}^{2+}$  inhibition which then permits activation of RyRs by ATP (Lamb *et al.*, 1991).

$\text{Mg}^{2+}$  is thought to inhibit RyRs by binding both to low affinity sites that show little specificity between divalent ions (I-sites) and to high affinity sites for  $\text{Ca}^{2+}$  (A-sites) thus preventing  $\text{Ca}^{2+}$  from activating the channel (Laver *et al.*, 1997). However, ATP is known to activate RyRs in the absence of cytoplasmic  $\text{Ca}^{2+}$  so it is not clear how  $\text{Mg}^{2+}$  at the A-sites affects channel opening under physiological conditions. Here we investigate the mechanism of  $\text{Mg}^{2+}$  inhibition in the presence of ATP and two drugs, 4,4'-diisothiocyano-stilbene-2,2'-disulfonic acid (DIDS) and ryanodine, which also activate RyRs in the absence of  $\text{Ca}^{2+}$ .

RyRs were isolated from rabbit skeletal muscle and incorporated into lipid bilayers using standard techniques (O'Neill *et al.*, 2003). Skeletal muscle was removed from dead rabbits. Cytoplasmic solutions contained 250 mM  $\text{Cs}^+$  (230 mM  $\text{CsCH}_3\text{O}_3\text{S}$  and 20 mM  $\text{CsCl}$ ) 10 mM TES at pH 7.4. Luminal solutions contained 50 mM  $\text{Cs}$  (30 mM  $\text{CsCH}_3\text{O}_3\text{S}$  and 20 mM  $\text{CsCl}$ ), 10 mM TES, pH 7.4.

DIDS decreased I-site affinity for  $\text{Mg}^{2+}$  and  $\text{Ca}^{2+}$  by 10 fold and ryanodine abolished binding completely. Cytoplasmic  $\text{Mg}^{2+}$  inhibited RyRs via the  $\text{Ca}^{2+}$  activation site even in the absence of  $\text{Ca}^{2+}$  indicating that  $\text{Mg}^{2+}$  inhibition is not merely due to the prevention of  $\text{Ca}^{2+}$  binding. In the case of ryanodine modified RyRs, monovalent ions ( $\text{Cs}^+$ ) could also activate the channel. RyR activity in the virtual absence of  $\text{Ca}^{2+}$  (~1 nM) was not due to sensitisation of the channel to  $\text{Ca}^{2+}$  as previously thought (Du *et al.*, 2001; Masumiya *et al.*, 2001) but was due to  $\text{Ca}^{2+}$ -independent channel opening by ryanodine. The apparent  $\text{Mg}^{2+}$  affinity at the A-site was decreased by cytoplasmic  $\text{Cs}^+$  and  $\text{Ca}^{2+}$  as well as by luminal  $\text{Ca}^{2+}$  in a way which suggests that cytoplasmic  $\text{Mg}^{2+}$ ,  $\text{Cs}^+$  and  $\text{Ca}^{2+}$  compete for a site near the cytoplasmic entrance. Ions at this site may progress to the A-site further into the pore. Binding of these ions at the A-site is in competition with luminal  $\text{Ca}^{2+}$  and leads to either activation ( $2 \times \text{Cs}^+$  or  $\text{Ca}^{2+}$ ) or inhibition ( $\text{Mg}^{2+}$ ) of RyRs.

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