## Phospholipase C $\gamma$ is essential for activation of store-operated Ca<sup>2+</sup> channels in liver cells

*T. Litjens*<sup>1</sup>, *T. Nguyen*<sup>1</sup>, *E. Aromataris*<sup>1</sup>, *M. Roberts*<sup>1</sup>, *G. Barritt*<sup>2</sup> and <u>G. Rychkov</u><sup>1</sup>, <sup>1</sup>School of Molecular and Biomedical Science, The University of Adelaide, Adelaide, SA 5005 and <sup>2</sup>School of Medicine, Flinders University of South Australia, G.P.O. Box 2100, Adelaide, SA 5001, Australia.

Release of  $Ca^{2+}$  from intracellular stores in non-excitable cells results in activation of  $Ca^{2+}$  influx through so-called store-operated  $Ca^{2+}$  channels (SOCs) on the plasma membrane (Putney *et al.*, 2001). Activation of these channels occurs in response to a decrease in the concentration of  $Ca^{2+}$  in the lumen of the endoplasmic reticulum, and it does not depend on how this decrease in  $[Ca^{2+}]$  is initiated. The molecular mechanism that underlies this phenomenon is poorly defined. Phospholipase  $C\gamma$  (PLC $\gamma$ ) has been previously shown to be either directly involved in activation of SOCs or to modulate their activity through the production of additional IP<sub>3</sub> in a number of cell lines (Patterson *et al.*, 2002). The identity of the SOCs regulated by PLC $\gamma$ , however, has not been established.

In this work we used short interfering RNA (siRNA) to specifically reduce the expression of the genes encoding PLC $\gamma$ 1 and PLC $\gamma$ 2 and whole cell patch clamping technique to measure activation of store-operated Ca<sup>2+</sup> current (I<sub>SOC</sub>) in H4IIE liver cells. Immunofluorescence and Western blotting were employed to verify the effectiveness of siRNA and the time course of the knock down of PLC $\gamma$ .

We have found that transfection of H4IIE liver cells with siRNA against PLC $\gamma$ 1 results in time dependent reduction of PLC $\gamma$ 1 protein with maximal effect apparent at 72-96 h. At the same time the amplitude of the I<sub>SOC</sub> developed in response to intracellular perfusion with IP<sub>3</sub> in cells transfected with siRNA against either PLC $\gamma$ 1 or 2 has decreased. The average maximal amplitude of I<sub>SOC</sub> decreased from -3.3±0.2 pA/pF (n=23) in control cells to -2.3±0.3 pA/pF (n=15) in cells transfected with siRNA against PLC $\gamma$ 1 and to -1.5±0.25 pA/pF (n=13) in cells transfected with siRNA against PLC $\gamma$ 2 together resulted in further reduction of the current to -0.65±0.17 pA/pF (n=14). Similar results were obtained when thapsigargin was used to activate I<sub>SOC</sub> instead of IP<sub>3</sub>. It is concluded that PLC $\gamma$  is required for activation of I<sub>SOC</sub> in liver cells, however, the catalytic activity of PLC $\gamma$  in this process in not essential.

Putney, J.W., Jr., Broad, L.M., Braun, F.J., Lievremont, J.P. & Bird, G.S. (2001) Journal of Cell Science 114, 2223-2229.

Patterson, R.L., van Rossum, D.B., Ford, D.L., Hurt, K.J., Bae, S.S., Suh, P.G., Kurosaki, T., Snyder, S.H. & Gill, D.L. (2002) Cell 111, 529-541.