

Rat skeletal muscle 3-O-MFPase activity is not decreased by fatiguing *in vitro* electrical stimulation

C.A. Goodman,^{1,3} A. Hayes^{2,3} and M.J. McKenna,^{1,3} ¹School of Human Movement, Recreation and Performance, Victoria University, Melbourne, VIC 8001, Australia, ²School of Biomedical and Health Sciences, Victoria University, Melbourne, VIC 8001, Australia and ³Centre for Ageing, Rehabilitation, Exercise and Sport, Victoria University, Melbourne, VIC 8001, Australia.

Maximal 3-O-MFPase activity is a surrogate measure of the Na⁺,K⁺-ATPase activity and is commonly utilized in human exercise studies involving skeletal muscle biopsy samples. Studies to date (e.g. Fowles *et al.*, 2002; Petersen *et al.*, 2005), using a range of exercise intensities and durations, have shown that an acute bout of fatiguing exercise results in a decrease in maximal 3-O-MFPase activity at the point fatigue of between ~11 to 38%. These observations have been interpreted as evidence that the maximal activity of the Na⁺,K⁺-ATPase has declined thus contributing to skeletal muscle fatigue. It is speculated that endogenous factors such as reactive oxygen species (ROS) and calcium (Ca²⁺) may be responsible for this observed reduction in 3-O-MFPase activity at fatigue. The aim of this study was to investigate whether rat skeletal muscle 3-O-MFPase activity could be reduced further than the above range from human studies by using large isolated skeletal muscles subjected to intense fatiguing *in vitro* electrical stimulation.

Sprague Dawley (260 ± 9g; Mean ± SE) fast twitch *extensor digitorum longus* (EDL) muscles (132 ± 4 mg) were dissected out under anaesthesia (Nembutal; ~85mg/kg) in accordance with Victoria University AEEC procedures and subjected to one of two different stimulation protocols: 1) two bouts of 10s continuous stimulation at a frequency of 100Hz (0.2ms pulse duration) separated by a 1hr recovery period; 2) two bouts of three min intermittent stimulation (1s stimulation at 100Hz followed by 4s recovery) separated by a 1hr recovery period. Tetanic force (500ms, 100Hz, 0.2ms pulse duration) was monitored during recovery. Fatigued muscles and their non-fatigued contra-lateral controls were blotted, weighed, frozen in liquid N₂ and maximal 3-O-MFPase activity analysed (Fraser & McKenna, 1998).

At the end of the first bout of 10s continuous stimulation tetanic force had declined by 51.8 ± 1.8% (*n* = 8) of initial force. Characteristic of high frequency fatigue, force had recovered to 81.2 ± 2.1% of initial after one min and remained relatively constant over the next hour (87.4 ± 2.6% of initial force at one hour). The second stimulation bout reduced force by 50.3 ± 1.3% of initial force, while 3-O-MFPase activity showed no decline (100.5 ± 3.4%; *p* = 0.9) compared to the non-fatigued, contra-lateral controls. Three minutes of high frequency intermittent stimulation resulted in tetanic force declining by 87.0 ± 1.0% (*n* = 8) of initial force. After one hour of recovery, tetanic force had gradually recovered to 62.7 ± 2.1% of initial force. At this time, force-frequency analysis showed the presence of low frequency fatigue with relative force being significantly lower at 10 (39.0 ± 2.1% vs 47.0 ± 1.2%; *p* = 0.005), 30 (44.4 ± 1.4% vs 60.7 ± 1.3%; *p* < 0.0001) and 50Hz (76.2 ± 1.4% vs 86.7 ± 0.7%; *p* < 0.0001) compared to pre-fatigue force. The second intermittent stimulation bout reduced force by 83.3 ± 1.3% of initial force while 3-O-MFPase activity was not significantly altered (94.4 ± 3.7%; *p* = 0.2) when compared to the non-fatigued contra-lateral controls.

In conclusion, under these conditions, rat EDL 3-O-MFPase activity was not reduced by either of the two fatiguing *in vitro* electrical stimulation protocols. Thus the decline in muscle force was not related to a depression in maximal 3-O-MFPase activity. Whether this reflects a species difference with resistance to Na⁺,K⁺-ATPase inactivation in the rat is unclear.

Fowles JR, Green HJ, Tupling R, O'Brien S & Roy BD. (2002) *Journal of Applied Physiology*, **92**: 1585-93.

Fraser SF & McKenna MJ. (1998) *Analytical Biochemistry* 258:63-7. (2006) *Journal of Physiology*, **576**: 279-88.

Petersen AC, Murphy KT, Snow RJ, Leppik JA, Aughey RJ, Garnham AP, Cameron-Smith D & McKenna MJ. (2005) *American Journal of Physiology*, **289**: R266-74.