

GABA_A receptors increase their conductance through novel protein interactions

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Native GABA_A channels display a single-channel conductance ranging between ~10-90 pS. Diazepam increases the conductance of some of these native channels but never those of recombinant receptors unless they are co-expressed with GABARAP. This trafficking protein clusters recombinant receptors in the membrane suggesting that high-conductance channels arise from receptors that are at locally high concentrations. The amphipathic (MA) helix that is present in the large cytoplasmic loop of every subunit of all ligand-gated ion channels mediates protein-protein interactions. Here we report that when applied to inside-out patches, a peptide mimicking the MA helix of the $\gamma 2$ subunit ($\gamma 381-403$) of the GABA_A receptor abrogates the potentiating effect of diazepam on both endogenous receptors and recombinant GABA_A receptors co-expressed with GABARAP, by substantially reducing their conductance. The protein interaction disrupted by the peptide did not involve GABARAP because a shorter peptide ($\gamma 386-403$) known to compete with the $\gamma 2$: GABARAP interaction did not affect the conductance of recombinant $\alpha\beta\gamma$ receptors co-expressed with GABARAP. The requirement for receptor clustering and the fact that the $\gamma 2$ MA helix is able to self-associate support a mechanism whereby adjacent GABA_A receptors interact *via* their $\gamma 2$ subunit MA helices, altering ion permeation through each channel. This finding has important implications for understanding both the structural design of ligand-gated ion channels and the adaptive, dynamic means a cell invokes to amplify its signalling capacity.