Dysfunctional intracellular calcium cycling in cardiac alternans

Joshua N. Edwards and Lothar A. Blatter

Department of Molecular Biophysics and Physiology, Rush University Medical Center, Chicago, IL 60612, USA.

Summary

- 1. Cardiac alternans refers to a condition in which there is a periodic beat-to-beat oscillation in electrical activity and the strength of cardiac muscle contraction at a constant heart rate. Clinically, cardiac alternans occurs in settings that are typical for cardiac arrhythmias and has been causally linked to these conditions.
- **2.** At the cellular level, alternans is defined as beat-to-beat alternations in contraction amplitude (mechanical alternans), action potential duration (APD; electrical or APD alternans), and Ca^{2+} transient amplitude (Ca^{2+} alternans).
- 3. The cause of alternans is multifactorial, however alternans always originate from disturbances of the bidirectional coupling between membrane voltage (V_m) and intracellular calcium ($[Ca^{2+}]_i$). Bi-directional coupling refers to the fact that in cardiac cells, V_m depolarization and the generation of action potentials cause the elevation of $[Ca^{2+}]_i$ that is required for contraction (a process referred to as excitation-contraction coupling). The changes of $[Ca^{2+}]_i$ on the other hand control V_m because important membrane currents are Ca^{2+} -dependent.
- **4.** Evidence is mounting that alternans is ultimately caused by disturbances of cellular Ca²⁺ homeostasis. Here we review how two key factors of cardiac cellular Ca²⁺ signaling the release of Ca²⁺ from internal stores and the capability of clearing the cytosol from Ca²⁺ after each beat determine the conditions under which alternans occurs. The contributions from key Ca²⁺ handling proteins surface membrane channels, ion pumps and transporters, and internal Ca²⁺ release channels are discussed.

Introduction

Cardiac alternans refers to a condition characterized by a periodic beat-to-beat oscillation in electrical activity and the strength of cardiac muscle contraction at a constant heart rate. The clinical manifestations of alternans occur in many settings in which arrhythmias are also common; however, its origin can be followed to the cellular and subcellular level. Here, we will review the alternans field from the perspective of the cellular disturbances of electrical and calcium signaling which lead to the proarrhythmic condition of alternans.

Excitation-contraction coupling in cardiac muscle

Each heartbeat requires a coordinated activation of cardiac muscle cells to sustain the pump function of the heart. Excitation-contraction coupling describes the process that converts electrical activation into mechanical activity and muscle contraction. The sequence of events begins with

depolarization of the surface membrane potential (V_m) by an action potential, followed by the entry of extracellular calcium through voltage-gated sarcolemmal L-type Ca²⁺ channels (also referred to as dihydropyridine receptors, DHPRs). Ca²⁺ influx triggers intracellular Ca²⁺ release by activating Ca²⁺-sensitive Ca²⁺ release channels (ryanodine receptors, RyRs) in the sarcoplasmic reticulum (SR) membrane by a mechanism termed Ca²⁺-induced Ca²⁺ release (CICR). The amplified Ca²⁺ release from the SR raises intracellular [Ca²⁺] ([Ca²⁺]_i) which activates the contractile apparatus and force is produced. Relaxation of cardiac cells is dependent upon mechanisms that lower [Ca²⁺], through reuptake into the SR Ca^{2+} sarcoplasmic/endoplasmic reticulum **ATPase** (SERCA) and extrusion from the cell primarily via sarcolemmal sodium-calcium exchange (NCX). Reuptake of Ca²⁺ provides the necessary filling of the SR to allow sufficient Ca²⁺ for release during the next heartbeat.

Ventricular myocytes typically have a well-developed transverse (t) tubular system. The t-tubular system consists of invagination of the surface membrane that extends as a 3-dimensional network of narrow transverse tubules throughout the longitudinal entire Approximately 30-50% of the sarcolemma exists as the ttubular system and forms a well-connected membrane network within the cell, but contiguous with the extracellular space. DHPRs together with many other ion channels and transporters are located in the t-tubular membrane. Clusters of RyRs on the terminal cisternae of the SR membrane appose DHPRs separated only by a narrow (a few nanometers) cleft, forming a dyad of two adjacent membranes.³ The dyad is the functional unit of SR Ca²⁺ release, termed SR Ca²⁺ release unit (CRU)⁴ or couplon.⁵ Ca²⁺ sparks are considered the elementary events of Ca²⁺ signaling in cardiac cells⁶ arising from CICR at individual CRUs, and according to the 'local control' model of cardiac excitation-contraction coupling⁷ are recruited independently and spatially summate to produce a Ca²⁺ transient.8,9 The well developed t-tubular network in ventricular myocytes ensures simultaneous activation of SR Ca²⁺ release throughout the entire ventricular myocyte during an action potential, resulting in spatially separate rather homogeneous Ca²⁺ transients (Figure 1A).

The fundamental process of excitation-contraction coupling in atrial and ventricular cells shows similarities, but also important structural and functional differences. The t-tubular system in atrial cells is significantly less developed or even entirely absent, 10,11 although there are species differences. For example, rudimentary t-tubular structures are found in rat, 12 sheep 13 and dog. 14 The spatial vicinity to the surface membrane defines two types of SR, termed

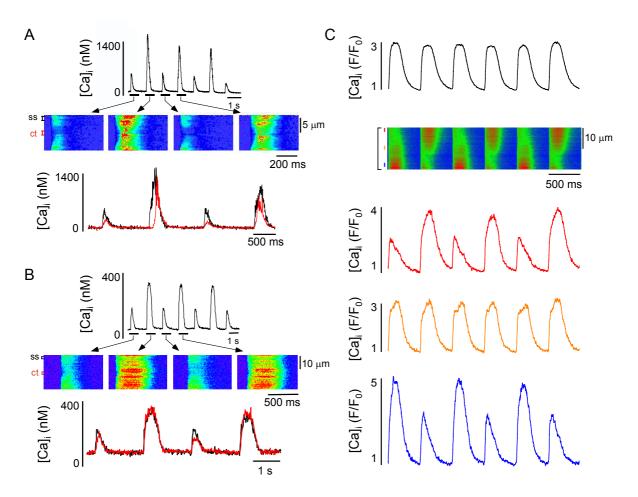


Figure 1. Cellular and subcellular Ca^{2+} alternans in cardiac myocytes. A,B: Spatiotemporal characteristics of Ca^{2+} transients during alternans in a atrial (A) and ventricular (B) myocyte. From top: whole cell Ca^{2+} transients, transverse confocal line scan images and subcellular $[Ca^{2+}]_i$ profiles recorded from subsarcolemmal (ss, black; corresponding to j-SR Ca^{2+} release) and central (ct, red; corresponding to nj-SR Ca^{2+} release) regions of the myocyte. Panels A and B modified from Hüser et al. with permission. C: Spatiotemporal characteristics of Ca^{2+} transients during alternans in an atrial myocyte where subcellular discordant or 'out-of-phase' alternans are present. The global $[Ca^{2+}]_i$ profile suggests no Ca^{2+} alternans, however spatially restricted profiles identify subcellular regions with no alternans coexisting with regions alternating out-of-phase.

junctional (j-SR) and non-junctional (nj-SR) SR. Because of the absence of t-tubules in atrial myocytes j-SR is restricted to the cell periphery. Both j-SR and nj-SR express RyRs, and - compared to ventricular myocytes - have a higher density of IP₃ receptors. 15,16 In atrial cells, peripheral j-SR and the more centrally located nj-SR are capable of active and robust SR Ca²⁺ release, however the mechanism of activation differs. Action potential-induced membrane depolarization activates Ca2+ entry through L-type Ca2+ channels which triggers CICR from RyRs of the j-SR. Elevation of peripheral [Ca²⁺], propagates then via CICR in a Ca²⁺ wave-like fashion in centripetal direction by a diffusion-reaction process or a 'fire-diffuse-fire' mechanism (Figure 1B). As a characteristic consequence of this mode of activation and ultrastructural arrangements, Ca²⁺ release is spatially inhomogeneous¹⁶⁻¹⁸ with complex subcellular [Ca²⁺], gradients (Figures 1B and 1C). These structural and functional differences are important for the susceptibility to spontaneous pro-arrhythmic Ca^{2+} release events (Ca^{2+} waves) and the propensity to develop cardiac alternans as will be discussed below.

Cardiac alternans

In 1872, for the first time a very interesting phenomenon, consisting of beat-to-beat oscillations in arterial pressure that occurred while the heart rate remained constant, was reported by Traube. 19 This observation, called 'pulsus alternans' would ultimately be known as mechanical alternans. With the arrival of electrocardiogram (ECG) similar beat-to-beat alternations of electrical activity of the heart (electrical alternans) were recorded in laboratory animals²⁰ and humans,²¹ and are typically referred to as repolarization or T-wave alternans. It was recognized early on that conditions of pulsus alternans were associated with severe cardiac pathologies and poor prognosis.²² To date, it is well established that cardiac

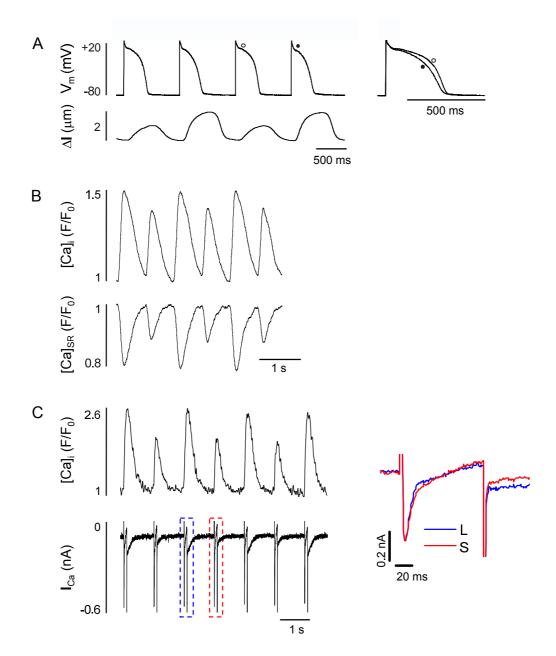


Figure 2. Electrical, mechanical and Ca^{2+} alternans in cardiac myocytes. A: Simultaneous recordings of action potentials and cell shortening from a single ventricular myocyte revealing discordant electromechanical alternans. To the right, two action potentials recorded during successive small- (open circle) and large-amplitude (filled circle) shortenings are superimposed to illustrate the differences in duration and kinetics. Modified from Hüser et al. With permission. B: Simultaneous recordings of cytosolic ($[Ca^{2+}]_i$; top) and intra-SR ($[Ca^{2+}]_{SR}$; bottom) Ca^{2+} alternans from a single ventricular myocyte. C: Simultaneous recordings of $[Ca^{2+}]_i$ (top) and I_{Ca} (bottom) in voltage-clamped atrial myocytes. To the right, an overlay of I_{Ca} measured during a large-amplitude Ca^{2+} transient (L; blue trace) and a small-amplitude Ca^{2+} transient (S; red trace) shows that Ca^{2+} alternans are not accompanied by alternating peak I_{Ca} . Modified from Shrkyl et al. with permission.

alternans is linked to increased risk for atrial and ventricular arrhythmias and sudden cardiac death across a wide range of pathophysiological conditions, including ischemia and myocardial infarction. Toward alternans in the ECG and microvolt electrical alternans testing have become a prognostic tool for arrhythmia risk stratification and antiarrhythmic therapy. 30-32

At the cellular level, cardiac alternans is defined by

cyclic, beat-to-beat alternations in contraction amplitude (mechanical alternans), action potential duration (APD; electrical or APD alternans), and Ca²⁺ transient amplitude (Ca²⁺ alternans) at constant stimulation frequency (Figure 2). Alternans is induced typically by rapid heart rates, however, the pacing threshold required to initiate it is influenced by a wide variety of factors and conditions³³⁻³⁶ and varies among different mammalian species.^{37,38}

Conditions that lower the pacing threshold include hypothermia, $^{38-42}$ interference with cellular energy metabolism through inhibition of glycolysis, $^{10,43-45}$ hypocalcaemia, 38,41,46,47 disturbance of mitochondrial functions, 44,48,49 hypercapnic acidosis, 50,51 ischemia, $^{52-56}$ hypertrophy, 57 IP $_3$ receptor-dependent Ca $^{2+}$ release, 58,59 and heart failure. $^{60-62}$ A shift to a higher pacing threshold for alternans has been reported in conditions of hypercalcaemia, 38,46 pharmacological sensitization of the SR Ca $^{2+}$ release channels, 63 and calcium channel antagonists. 42,54 Interestingly, β -adrenergic stimulation, while generally having positive inotropic effects, can either enhance 64 or suppress 10,44,48 alternans (cf. discussion below).

Mechanism of cardiac alternans: bi-directional coupling between V_m and $\left[Ca^{2+}\right]_i$

The plethora of studies on cardiac alternans clearly document that this proarrhythmic condition multifactorial. Nonetheless, it is generally agreed that instabilities of the bi-directional coupling of V_m and [Ca²⁺]_i are a cucial factor for the generation of alternans. 'Bidirectional' coupling refers to the fact that membrane depolarization in form of an action potential is required to initiate Ca2+ release and to elevate [Ca2+], however the ensuing dynamics of $\left[Ca^{2+}\right]_i$ affect V_m through the Ca²⁺-dependence of numerous membrane conductances as outlined below.65 Consequently, the question arises as to whether alternans are either V_m or $[Ca^{2+}]_i$ driven. 35,36,66,67 As such, a classic 'chicken or egg conundrum' exists in the literature relating to the fact that the mechanisms responsible alternans remain incompletely for understood.68-70

$$V_m \rightarrow [Ca^{2+}]_i$$
 coupling

V_m-driven alternans is determined by a single parameter - APD restitution. The key concept behind the paradigm of V_m -driven alternans is that APD restitution is a time-dependent process resulting from the fact that recovery from inactivation of ion currents underlying the action potential requires time (thus resulting in absolute and relative refractoriness of excitability). APD restitution is defined as the relationship between APD and diastolic interval (DI). The heart rate is inversely related to cycle length (CL), which is calculated as CL = APD + DI. When heart rate increases, the APD shortens to preserve the diastolic interval for ventricular filling. Therefore, electrical alternans is critically dependent on beat-to-beat changes in diastolic interval. $V_m \rightarrow [Ca^{2+}]_i$ coupling is generally believed to be positive, i.e., a long APD is paralleled by a strong contraction and large amplitude Ca²⁺ transient. Positive coupling between APD and Ca2+ transient or contraction amplitude is also referred to as 'in-phase' or 'concordant' at the cellular level. 'Negative' $V_m \leftrightarrow [Ca^{2+}]_i$ coupling results in 'discordant' or 'out-of-phase' alternans at the single cell level (Figure 2A). The term 'discordant' is also used at the multicellular tissue level where it refers to different regions of the myocardium alternating asynchronously or 'out-of-phase'. Such regions are separated by nodal lines 71 which mark areas of highest $\left[\text{Ca}^{2+}\right]_{i}$ and APD gradients and become sites of origin for arrhythmias. The terminology discordant/concordant is also used at the subcellular level and describes alternans pattern of subcellular regions within a single cell (Figure 1C). 43,72,73 Alternations of the diastolic interval is critical for the availability of the L-type Ca^{2+} channel current ($\text{I}_{\text{Ca},\text{L}}$) at a given heartbeat. A longer diastolic interval allows more time for recovery of $\text{I}_{\text{Ca},\text{L}}$, leading to enhanced $\text{I}_{\text{Ca},\text{L}}$, larger Ca^{2+} release and longer APD during the following beat. Now the longer APD is followed by a shorter diastolic interval, leading to less recovery of $\text{I}_{\text{Ca},\text{L}}$ with less Ca^{2+} release and shorter APD during the next beat, thus sustaining alternans.

$$[Ca^{2+}]_i \rightarrow V_m$$
 coupling

 $[Ca^{2+}]_i{\to}V_m$ coupling is determined by the fact that $[Ca^{2+}]_i$ feeds back on V_m . This occurs through the Ca^{2+} -dependence of ion channels and transporters, $\emph{i.e.}$ membrane conductances that in turn also control $[Ca^{2+}]_i$ cycling. With respect to cardiac alternans, $I_{Ca,L}$ and I_{NCX} are most important. 35,36 $[Ca^{2+}]_i{\to}V_m$ coupling can be positive or negative depending on which of the Ca^{2+} -dependent ion currents or transporters dominates. For example, a positive $[Ca^{2+}]_i{\to}V_m$ coupling occurs when the large Ca^{2+} transient causes a prolongation of APD by potentiating the inward I_{NCX} (1 Ca^{2+} ion extruded in exchange to 3 Na^+ ions) to a greater extent than reducing $I_{Ca,L}$ through Ca^{2+} -dependent inactivation.

Negative $[Ca^{2+}]_i \rightarrow V_m$ coupling occurs when reduction of $I_{Ca,L}$ dominates over increased I_{NCX} which ultimately results in APD shortening. Other Ca^{2+} -sensitive currents (non-selective cation current, Cl^- current) may modulate $[Ca^{2+}]_i \rightarrow V_m$ coupling, but appear to be quantitatively less important.

Two key parameters relevant to the generation of [Ca²⁺];-driven alternans at the cellular level are i) fractional Ca²⁺ release from the SR and SR Ca²⁺ load, and ii) the efficiency of beat-to-beat cytosolic Ca²⁺ sequestration.^{35,36} Fractional release of Ca²⁺ refers to the nonlinear relationship between the end-diastolic SR Ca²⁺ content and the amount of Ca²⁺ (or % of SR Ca²⁺ content) released by CICR with each heartbeat (i.e., a larger fraction of Ca²⁺ is released at a higher SR Ca²⁺ content).⁷⁴ Ca²⁺ sequestration is a phenomenological parameter and refers to the net efficiency of cytosolic Ca²⁺ removal. It is dependent on i) the activity of SERCA to reload the SR, ii) Na⁺/Ca²⁺ exchange and plasmalemmal Ca2+-ATPase activity to extrude Ca²⁺ from the cell, iii) cytosolic buffering (including mitochondrial Ca²⁺ uptake), and iv) diastolic SR Ca²⁺ leak. Therefore, alternans can occur at modest SR loads and small fractional releases under conditions where Ca²⁺ sequestration is low. Alternatively at high sequestration rates, higher Ca²⁺ loads and fractional release are required to induce alternans. In general, factors increasing fractional release promote, and factors increasing Ca²⁺ sequestration efficiency protect against alternans. To illustrate, in heart failure where SERCA expression is reduced and Ca^{2+} release from the SR is increased, or during acute cardiac ischemia (where SR Ca^{2+} load is initially unaffected, but SERCA activity is diminished due to reduced ATP levels), the heart is pushed into instability due to diminished Ca^{2+} sequestration. On the other hand, under β -adrenergic stimulation SERCA activity and consequently SR Ca^{2+} uptake and load are increased, leading to enhanced fractional release that tends to promote alternans. Increased SERCA activity, however, also increases the efficiency of Ca^{2+} sequestration, resulting in protection against alternans. Whether β -adrenergic stimulation favors or protects α or protects α against alternans and alternans-related arrhythmias depends on which α -adrenergic effects predominate.

Recently, an overarching conceptual model for cardiac alternans has been forwarded, termed '3R theory'. 34,75,76 The 3R theory links Ca^{2+} spark properties, i.e. the properties of Ca²⁺ release from individual CRUs, to whole-cell Ca2+ alternans. Ca2+ alternans occurs due to instabilities of the relationship of 3 critical spark properties (the '3 Rs'): 1) Randomness of Ca²⁺ sparks, 2) Recruitment of sparks by neighboring CRUs, and 3) Refractoriness of a CRU. An individual CRU can be in 3 different states: recovered (i.e. ready to fire), firing or refractory. The theory predicts (by numerical computations) that alternans occurs when the probability of a spontaneous primary spark is intermediate (intermediate randomness), coupling among CRUs is strong (high probability of a primary spark triggering a secondary spark from a neighboring CRU; high degree of recruitment), and a high degree of refractoriness is prevalent (i.e. the probability of a CRU not being recovered from previous release is high). This unifying theoretical framework predicts how Ca²⁺ cycling proteins and organelles (L-type Ca2+ channels, RyR, SERCA, NCX, Ca²⁺ buffers and mitochondria) affect the 3 Rs and SR Ca²⁺ load, and thus the prevalence of Ca²⁺ alternans. Interestingly, in the 3R framework SR Ca²⁺ load is not an explicit parameter which is consistent with our observation that Ca2+ alternans are not dependent on alternating enddiastolic [Ca²⁺]_{SR}. 10,63,77 Nonetheless, SR Ca²⁺ load is a critical factor for Ca²⁺ alternans since load determines the efficiency of the L-type Ca²⁺ current to trigger release; it controls RyR function through its luminal Ca²⁺ sensitivity and influences refractoriness of release. In the next section we will summarize the specific contributions of the major Ca²⁺ signaling proteins and organelles to alternans.

As mentioned earlier, alternans is a recognized risk factor for ventricular and atrial arrhythmias. 70,78,79 [Ca²⁺]_i \rightarrow V_m coupling can be positive or negative, *i.e.*, result in both concordant and discordant alternans. At the level of the heart, spatially discordant alternans favor re-entry, triggering ectopic beats and facilitating the onset of lethal arrhythmic events 80,81 whereas concordant alternans is considered less arrhythmogenic. 82 At the cellular level atrial myocytes are particularly susceptible to Ca²⁺ alternans induced by pacing or metabolic inhibition. In atrial myocytes alternans is typically subcellularly inhomogeneous (Figures 1B and 1C). Subcellular

inhomogeneities consist of subcellular transverse and longitudinal gradients of the degree of Ca²⁺ alternans, and subcellular regions alternating out-of-phase. 10,16,43,45,59 These [Ca²⁺], gradients and inhomogeneities result from the unique structural and functional features of atrial excitationcontraction coupling and are consistent with simulation studies on the relationship between the lack of t-tubules and generation of alternans.83 We demonstrated that the complex subcellular [Ca²⁺], inhomogeneities of atrial alternans generates a substrate for spontaneous (i.e., not electrically triggered) proarrhythmic Ca2+ release and represents a mechanistic link to atrial arrhythmia at the cellular level.⁴³ Of particular interest is the observation of subcellular 'discordant' Ca2+ alternans where subcellular regions alternate out-of-phase (Figure 1C). subcellular areas are typically separated by regions where spontaneous Ca²⁺ waves originate with high probability, reminiscent of the nodal lines observed at tissue level.⁷¹

Thus, it appears that spatially discordant alternans phenomena at tissue level can be recapitulated at the cellular level.

Ca²⁺ handling proteins and organelles and their role in cardiac alternans

Although clearly a multifactorial phenomenon, consensus is emerging that electromechanical and Ca^{2+} alternans are ultimately linked to impaired $[Ca^{2+}]_i$ regulation, and $[Ca^{2+}]_i {\rightarrow} V_m$ coupling dominates the mechanisms that are responsible for the occurrence of alternans. $^{68,84-86}$ In the following paragraphs we will address the contributions of L-type Ca^{2+} channels, SR and Ca^{2+} load, the SR Ca^{2+} release machinery (RyRs) and mitochondria to alternans.

L-type Ca²⁺ channels

Considering that $I_{\text{Ca},L}$ is the critical trigger for CICR during excitation-contraction coupling and SR Ca^{2+} release is graded with the magnitude of the current, 87,88 beat-tobeat alternation of I_{Ca.L.} has been considered a candidate to cause alternans. A potential mechanism entails incomplete time-dependent recovery from inactivation of $I_{Ca,L}$ which could lead to Ca²⁺ alternans.⁸⁹⁻⁹¹ This hypothesis, however would have to reconcile the observation in both atrial (Figure 2C) and ventricular myocytes that alternans can occur while peak $I_{Ca,L}$ remains unchanged from one beat to the next. $^{10,63,77,92-94}$ Furthermore, mechanical and Ca^{2+} alternans can occur in the absence of APD alternans (confirmed in patch-clamp experiments) and with constant $I_{\text{Ca.L}}$. 10,63,92,95,96 Ca²⁺ alternans is observed even in myocytes stimulated at a high frequency during action potential voltage clamp in the absence of APD alternans.95 Together these data suggest that I_{Ca.L.} is unlikely paramount in the onset of alternans.

SERCA and SR Ca²⁺ load

It can be speculated that at higher pacing frequencies, limitations of SR Ca²⁺ uptake kinetics preclude adequate

refilling of Ca²⁺ stores, particularly after a large Ca²⁺ transient. Consequently, the reduced filling only permits a small Ca²⁺ transient during the next beat thus resulting in Ca²⁺ alternans. This led to the suggestion that beat-to-beat alternations in end-diastolic SR Ca²⁺ load is a prerequisite for alternans, ⁹³ possibly due to an instability in the feedback control of SR Ca²⁺ content. ⁹⁷ However, direct and dynamic measurements of intra-SR [Ca²⁺] have shown (Figure 2B) that alternans do not require beat-to-beat alternations in SR Ca²⁺ content. ^{10,63,77} The role of Ca²⁺ reuptake into the SR and reestablishing Ca²⁺ load has been further investigated by enhancing SERCA activity. ⁹⁸⁻¹⁰⁰ Indeed, using genetic approaches to up-regulate SERCA2a (cardiac isoform) resulted in suppression of alternans. ^{83,100-102}

RyR and restitution of SR Ca²⁺ release

The magnitude of a Ca²⁺ transient is determined by the recovery of the trigger of CICR (I_{Ca I} restitution), SR Ca²⁺ load and the release mechanism itself (RyRs and associated regulatory proteins) from the preceding heartbeat. If recovery of any of these parameters is incomplete, the subsequent Ca²⁺ transient is expected to be reduced, thus facilitating the onset of alternans. Ca²⁺ release is unavailable immediately after release due to RyR inactivation. Recovery of elementary Ca2+ sparks and whole-cell Ca²⁺ transients after a preceding release requires several hundred milliseconds to reach full availability. 103-108 Incomplete RyR recovery from inactivation may contribute to instabilities of Ca²⁺ release and vulnerability to alternans and arrhythmias, particularly when pacing frequencies overlap with the time scale of RyR and Ca²⁺ release restitution. 109 Thus, refractoriness of release and its timedependent recovery can become the critical factor for the occurrence of Ca2+ alternans, as has been shown experimentally¹¹⁰ and in computational studies.¹¹¹ In a comprehensive investigation we recently demonstrated refractoriness of SR Ca²⁺ release as the key causative factor for alternans in atrial tissue. Restitution properties and refractoriness of Ca²⁺ release during alternans were evaluated by four different approaches: 1) latency of spontaneous global Ca²⁺ releases (Ca²⁺ waves) and 2) Ca²⁺ spark frequency during rest after a large and a small alternans Ca2+ transient, 3) premature action potentialinduced Ca2+ transients after a large and a small beat, and 4) the efficacy of a photolytically induced Ca²⁺ signal to trigger additional Ca²⁺ release during alternans. The results showed that restitution of SR Ca²⁺ release was significantly delayed after the large Ca2+ transient, leading to the conclusion that beat-to-beat alternation of the timedependent restitution properties and refractory kinetics of SR Ca²⁺ release represents a key mechanism underlying alternans.63

Mitochondria

Mitochondria contribute to cardiac Ca^{2+} cycling and excitation-contraction coupling at different levels: as a major source of ATP (energetics) that provides the fuel for the contractile apparatus, sustains ion pumps and alters the

activity of Ca²⁺ handling proteins, for example through phosphorylation or acting as a direct modulator (e.g., modulation of RyR activity by MgATP). Mitochondria shape cytosolic Ca²⁺ signals directly through Ca²⁺ sequestration. Furthermore, mitochondria can be a major source of reactive oxygen species (ROS), thus determine the cellular redox environment which profoundly affects cardiac excitability and the activity of Ca²⁺ handling proteins, including the RyR and SERCA (for review, see Zima & Blatter, 2006¹¹²). The pivotal role of mitochondria for Ca²⁺ signaling and excitation-contraction coupling is further underscored by the fact that these organelles occupy approximately 35% of the cell volume. Despite the undisputed importance of mitochondria for cardiac Ca²⁺ signaling and excitation-contraction coupling, it is rather surprising that mitochondria have been rarely the topic of studies on alternans mechanism. 113 In two recent studies we demonstrated that impairment of mitochondrial functions enhanced alternans. 44,48 In these studies the application of pharmacological blockers targeted to the various mitochondrial functions all enhanced the degree of Ca²⁺ alternans induced by pacing. This could be achieved by dissipation of mitochondrial membrane potential, as well as by inhibition of mitochondrial F₂/F₂-ATP synthase, inhibition of electron transport chain and Ca-dependent dehydrogenases, and by blockage of mitochondrial Ca²⁺ uptake or extrusion. These results are in agreement with other studies that confirmed that mitochondrial uncoupling facilitates alternans, 49 and demonstrated that an altered redox environment can generate conditions that favor alternans.94 Thus, with all likelihood mitochondria will emerge as a critical factor for the development of alternans.

Concluding remarks

Cardiac alternans is an intriguing phenomenon with clinical implications to a range of cardiac pathologies, while also providing insights into the intricacies of cellular Ca²⁺ cycling in heart muscle. Although clearly a multifactorial process, the experimental, theoretical and computational data exploring electrical, Ca²⁺ and mechanical alternans indicate that dysfunctional Ca²⁺ cycling appears to be the crucial mechanistic link between the contractile dysfunction and electrical instabilities seen at the cellular level, as well as clinically in patients. Despite the complexity of cardiac Ca²⁺ signaling, recent years have seen remarkable progress towards the understanding of the phenomenon of cardiac alternans. Growing theoretical and experimental evidence emphasizes that cellular Ca²⁺ signaling - and particularly the key proteins responsible for beat-to-beat Ca²⁺ release - are at the 'heart' of the problem of cardiac alternans. The recognition of the central role of the cardiac Ca²⁺ release machinery for alternans will pave the way, by pharmacologically or genetically targeting these Ca²⁺ handling proteins, to develop novel therapeutic strategies for the suppression of cardiac arrhythmias.

Acknowledgments

This work was supported by an Early Career Fellowship from the National Health and Medical Research Council of Australia (JNE); and by National Institutes of Health Grants HL62231, HL80101 and HL101235, and the Leducq Foundation (LAB).

References

- 1. Fabiato A. Calcium-induced release of calcium from the cardiac sarcoplasmic reticulum. *Am. J. Physiol.* 1983; **245**: C1-14.
- Soeller C, Cannell MB. Examination of the transverse tubular system in living cardiac rat myocytes by 2-photon microscopy and digital image-processing techniques. Circ. Res. 1999; 84: 266-75.
- Flucher BE, Franzini-Armstrong C. Formation of junctions involved in excitation-contraction coupling in skeletal and cardiac muscle. *Proc. Natl. Acad. Sci. USA* 1996; 93: 8101-6.
- 4. Franzini-Armstrong C, Jorgensen AO. Structure and development of E-C coupling units in skeletal muscle. *Annu. Rev. Physiol.* 1994; **56**: 509-34.
- Stern MD, Song LS, Cheng H, Sham JS, Yang HT, Boheler KR, R[Unicode char 0355]Ãos E. Local control models of cardiac excitation-contraction coupling. A possible role for allosteric interactions between ryanodine receptors. *J. Gen. Physiol.* 1999; 113: 469-89.
- Cheng H, Lederer WJ, Cannell MB. Calcium sparks: elementary events underlying excitation-contraction coupling in heart muscle. *Science* 1993; 262: 740-4.
- 7. Stern MD. Theory of excitation-contraction coupling in cardiac muscle. *Biophys J* 1992; **63**:497-517.
- 8. Cannell MB, Cheng H, Lederer WJ. Spatial non-uniformities in $[Ca^{2+}]_i$ during excitation-contraction coupling in cardiac myocytes. *Biophys. J.* 1994; **67**:1942-56.
- 9. Cannell MB, Cheng H, Lederer WJ. The control of calcium release in heart muscle. *Science* 1995; **268**: 1045-9.
- 10. Huser J, Wang YG, Sheehan KA, Cifuentes F, Lipsius SL, Blatter LA. Functional coupling between glycolysis and excitation-contraction coupling underlies alternans in cat heart cells. *J. Physiol.* 2000; **524**: 795-806.
- 11. Berlin JR. Spatiotemporal changes of Ca²⁺ during electrically evoked contractions in atrial and ventricular cells. *Am. J. Physiol.* 1995; **269**: H1165-70.
- 12. Kirk MM, Izu LT, Chen-Izu Y, McCulle SL, Wier WG, Balke CW, Shorofsky SR. Role of the transverse-axial tubule system in generating calcium sparks and calcium transients in rat atrial myocytes. *J. Physiol.* 2003; **547**: 441-51.
- 13. Dibb KM, Clarke JD, Horn MA, Richards MA, Graham HK, Eisner DA, Trafford AW. Characterization of an extensive transverse tubular network in sheep atrial myocytes and its depletion in

- heart failure. Circ. Heart Fail. 2009; 2: 482-9.
- 14. Wakili R, Yeh YH, Yan Qi X, *et al.* Multiple potential molecular contributors to atrial hypocontractility caused by atrial tachycardia remodeling in dogs. *Circ. Arrhythm. Electrophysiol.* 2010; **3**: 530-41.
- 15. Greiser M, Lederer WJ, Schotten U. Alterations of atrial Ca²⁺ handling as cause and consequence of atrial fibrillation. *Cardiovasc. Res.* 2011; **89**: 722-33.
- Blatter LA, Kockskamper J, Sheehan KA, Zima AV, Huser J, Lipsius SL. Local calcium gradients during excitation-contraction coupling and alternans in atrial myocytes. J. Physiol. 2003; 546: 19-31.
- 17. Huser J, Lipsius SL, Blatter LA. Calcium gradients during excitation-contraction coupling in cat atrial myocytes. *J. Physiol.* 1996; **494**: 641-51.
- 18. Bootman MD, Smyrnias I, Thul R, Coombes S, Roderick HL. Atrial cardiomyocyte calcium signalling. *Biochim. Biophys. Acta* 2011; **1813**: 922-34.
- 19. Traube L. Ein Fall von Pulsus Bigeminus nebst Bemerkungen uber die Leberschwellungen bei Klappenfehlern and uber acute Leberatrophie. Berlin Klin. Wochenschr. 1872; 9: 185-8.
- 20. Hering HE. Experimentelle studien an Saugertherien uber das elektrocardiogramm. *Z. Exp. Pathol. Ther.* 1910; **7**: 363-78.
- 21. Lewis T. Notes upon alternation of the heart. *Quart. J. Med.* 1910; **4**: 141–4.
- 22. Windle JD. The incidence and prognostic value of the pulsus alternans in myocardial and arterial disease. *Quart. J. Med* 1913; **6**: 453–62.
- 23. Armoundas AA, Tomaselli GF, Esperer HD. Pathophysiological basis and clinical application of T-wave alternans. *J. Am. Coll. Cardiol.* 2002; **40**: 207-17.
- 24. Armoundas AA, Hohnloser SH, Ikeda T, Cohen RJ. Can microvolt T-wave alternans testing reduce unnecessary defibrillator implantation? *Nat. Clin. Pract. Cardiovasc. Med.* 2005; **2**: 522-8.
- 25. Dumitrescu C, Narayan P, Efimov IR, Cheng Y, Radin MJ, McCune SA, Altschuld RA. Mechanical alternans and restitution in failing SHHF rat left ventricles. *Am. J. Physiol. Heart Circ. Physiol.* 2002; **282**: H1320-6.
- 26. Merchant FM, Armoundas AA. Role of substrate and triggers in the genesis of cardiac alternans, from the myocyte to the whole heart: implications for therapy. *Circulation* 2012; **125**: 539-49.
- 27. Dilly SG, Lab MJ. Electrophysiological alternans and restitution during acute regional ischaemia in myocardium of anaesthetized pig. *J. Physiol.* 1988; **402**: 315-33.
- 28. Smith JM, Clancy EA, Valeri CR, Ruskin JN, Cohen RJ. Electrical alternans and cardiac electrical instability. *Circulation* 1988; 77: 110-21.
- 29. Verrier RL, Nearing BD. Electrophysiologic basis for T wave alternans as an index of vulnerability to ventricular fibrillation. *J. Cardiovasc.*

- Electrophysiol. 1994; 5: 445-61.
- 30. Rosenbaum DS, Jackson LE, Smith JM, Garan H, Ruskin JN, Cohen RJ. Electrical alternans and vulnerability to ventricular arrhythmias. *N. Engl. J. Med.* 1994; **330**: 235-41.
- 31. Verrier RL, Nieminen T. T-wave alternans as a therapeutic marker for antiarrhythmic agents. *J. Cardiovasc. Pharmacol.* 2010; **55**: 544-54.
- 32. Verrier RL, Klingenheben T, Malik M, El-Sherif N, Exner DV, Hohnloser SH, Ikeda T, Mart[Unicode char 0355]Ãnez JP, Narayan SM, Nieminen T, Rosenbaum DS. Microvolt T-wave alternans testing has a role in arrhythmia risk stratification. *J. Am. Coll. Cardiol.* 2012; **59**: 1572-3.
- 33. Euler DE. Cardiac alternans: mechanisms and pathophysiological significance. *Cardiovasc. Res.* 1999; **42**: 583-90.
- 34. Qu Z, Nivala M, Weiss JN. Calcium alternans in cardiac myocytes: order from disorder. *J. Mol. Cell Cardiol.* 2013; **58**: 100-9.
- 35. Weiss JN, Karma A, Shiferaw Y, Chen PS, Garfinkel A, Qu Z. From pulsus to pulseless: the saga of cardiac alternans. *Circ. Res.* 2006; **98**: 1244-53.
- 36. Weiss JN, Nivala M, Garfinkel A, Qu Z. Alternans and arrhythmias: from cell to heart. *Circ. Res.* 2011; **108**: 98-112.
- 37. Spear JF, Moore EN. A comparison of alternation in myocardial action potentials and contractility. *Am. J. Physiol.* 1971; **220**: 1708-16.
- 38. Lu HH, Lange G, Brooks CM. Comparative studies of electrical and mechanical alternation in heart cells. *J. Electrocardiol.* 1968; **1**: 7-17.
- Gilbert JL, Janse MJ, Lu HH, Pinkston JO, Brooks CM. Production and abolition of alternation in mechanical action of the ventricle. *Am. J. Physiol.* 1965; 209: 945-50.
- 40. Floyd WL, Dillon ML. Observations on sustained pulsus alternans during hypothermia. *Am. Heart J.* 1967; **73**: 765-76.
- 41. Wohlfart B. Analysis of mechanical alternans in rabbit papillary muscle. *Acta Physiol. Scand.* 1982; **115**: 405-14.
- Hirayama Y, Saitoh H, Atarashi H, Hayakawa H. Electrical and mechanical alternans in canine myocardium in vivo. Dependence on intracellular calcium cycling. *Circulation* 1993; 88: 2894-902.
- 43. Kockskamper J, Blatter LA. Subcellular Ca²⁺ alternans represents a novel mechanism for the generation of arrhythmogenic Ca²⁺ waves in cat atrial myocytes. *J. Physiol.* 2002; **545**: 65-79.
- 44. Florea SM, Blatter LA. The role of mitochondria for the regulation of cardiac alternans. *Front. Physiol.* 2010; **1**: 141.
- 45. Kockskamper J, Zima AV, Blatter LA. Modulation of sarcoplasmic reticulum Ca²⁺ release by glycolysis in cat atrial myocytes. *J. Physiol.* 2005; **564**: 697-714.
- 46. Badeer HS, Ryo UY, Gassner WF, Kass EJ, Cavaluzzi J, Gilbert JL, Brooks CM. Factors affecting pulsus alternans in the rapidly driven heart and papillary

- muscle. Am. J. Physiol. 1967; 213: 1095-101.
- 47. Surawicz B. Effect of Ca on Duration of Q-T Interval and Ventricular Systole in Dog. *Am. J. Physiol.* 1963; **205**: 785-9.
- 48. Florea SM, Blatter LA. Regulation of cardiac alternans by β-adrenergic signaling pathways. *Am. J. Physiol. Heart Circ. Physiol.* 2012; **303**: H1047-56.
- 49. Smith RM, Visweswaran R, Talkachova I, Wothe JK, Tolkacheva EG. Uncoupling the mitochondria facilitates alternans formation in the isolated rabbit heart. *Am. J. Physiol. Heart Circ. Physiol.* 2013; **305:** H9-18.
- 50. Lab MJ, Lee JA. Changes in intracellular calcium during mechanical alternans in isolated ferret ventricular muscle. *Circ. Res.* 1990; **66**: 585-95.
- 51. Orchard CH, McCall E, Kirby MS, Boyett MR. Mechanical alternans during acidosis in ferret heart muscle. *Circ. Res.* 1991; **68**: 69-76.
- 52. Parmley WW, Tomoda H, Fujimura S, Matloff JM. Relation between pulsus alternans and transient occlusion of the left anterior descending coronary artery. *Cardiovasc. Res.* 1972; **6**: 709-15.
- 53. Weber KT, Janicki JS, Fishman AP. Aerobic limit of the heart perfused at constant pressure. *Am. J. Physiol.* 1980; **238**: H118-25.
- 54. Hashimoto H, Suzuki K, Miyake S, Nakashima M. Effects of calcium antagonists on the electrical alternans of the ST segment and on associated mechanical alternans during acute coronary occlusion in dogs. *Circulation* 1983; **68**: 667-72.
- 55. Uno K. Mechanisms of pulsus alternans: its relation to alternation of regional contraction and elevated ST segment. *Am. Heart J.* 1991; **122**: 1694-700.
- 56. Murphy CF, Lab MJ, Horner SM, Dick DJ, Harrison FG. Regional electromechanical alternans in anesthetized pig hearts: modulation by mechanoelectric feedback. *Am. J. Physiol.* 1994; 267: H1726-35.
- 57. Kotsanas G, Holroyd SM, Young R, Gibbs CL. Mechanisms contributing to pulsus alternans in pressure-overload cardiac hypertrophy. *Am. J. Physiol.* 1996; **271**: H2490-500.
- 58. Shkryl VM, Maxwell JT, Blatter LA. A novel method for spatially complex diffraction-limited photoactivation and photobleaching in living cells. *J. Physiol.* 2012; **590**: 1093-100.
- 59. Zima AV, Blatter LA. Inositol-1,4,5-trisphosphate-dependent Ca²⁺ signalling in cat atrial excitation-contraction coupling and arrhythmias. *J. Physiol.* 2004; **555**: 607-15.
- 60. Brooks WW, Bing OH, Litwin SE, Conrad CH, Morgan JP. Effects of treppe and calcium on intracellular calcium and function in the failing heart from the spontaneously hypertensive rat. *Hypertension* 1994; **24**: 347-56.
- 61. Narayan P, McCune SA, Robitaille PM, Hohl CM, Altschuld RA. Mechanical alternans and the force-frequency relationship in failing rat hearts. *J. Mol. Cell Cardiol.* 1995; **27**: 523-30.

- 62. Wilson LD, Jeyaraj D, Wan X, Hoeker GS, Said TH, Gittinger M, Laurita KR, Rosenbaum DS. Heart failure enhances susceptibility to arrhythmogenic cardiac alternans. *Heart Rhythm* 2009; **6**: 251-9.
- 63. Shkryl VM, Maxwell JT, Domeier TL, Blatter LA. Refractoriness of sarcoplasmic reticulum Ca²⁺ release determines Ca²⁺ alternans in atrial myocytes. *Am. J. Physiol. Heart Circ. Physiol.* 2012; **302**: H2310-20.
- 64. de Diego C, Chen F, Xie LH, Dave AS, Thu M, Rongey C, Weiss JN, Valderrábano M. Cardiac alternans in embryonic mouse ventricles. *Am. J. Physiol Heart Circ. Physiol.* 2008; **294**: H433-40.
- Shiferaw Y, Sato D, Karma A. Coupled dynamics of voltage and calcium in paced cardiac cells. *Phys. Rev. E. Stat. Nonlin. Soft Matter Phys.* 2005; 71: 021903.
- 66. Jordan PN, Christini DJ. Characterizing the contribution of voltage-and calcium-dependent coupling to action potential stability: implications for repolarization alternans. *Am. J. Physiol. Heart Circ. Physiol.* 2007; 293: H2109-18.
- 67. Sato D, Shiferaw Y, Garfinkel A, Weiss JN, Qu Z, Karma A. Spatially discordant alternans in cardiac tissue: role of calcium cycling. *Circ. Res.* 2006; **99**: 520-7.
- 68. Eisner DA, Li Y, O'Neill SC. Alternans of intracellular calcium: mechanism and significance. *Heart Rhythm* 2006; **3**: 743-5.
- 69. Qu Z, Weiss JN. The chicken or the egg? Voltage and calcium dynamics in the heart. *Am. J. Physiol. Heart. Circ. Physiol.* 2007; **293**: H2054-5.
- Walker ML, Rosenbaum DS. Repolarization alternans: implications for the mechanism and prevention of sudden cardiac death. *Cardiovasc. Res.* 2003; 57: 599-614.
- 71. Hayashi H, Shiferaw Y, Sato D, Nihei M, Lin SF, Chen PS, Garfinkel A, Weiss JN, Qu Z. Dynamic origin of spatially discordant alternans in cardiac tissue. *Biophys. J.* 2007; **92**: 448-60.
- Cordeiro JM, Malone JE, Di Diego JM, Scornik FS, Aistrup GL, Antzelevitch C, Wasserstrom JA. Cellular and subcellular alternans in the canine left ventricle. Am. J. Physiol. Heart Circ. Physiol. 2007; 293: H3506-16.
- 73. Aistrup GL, Kelly JE, Kapur S, Kowalczyk M, Sysman-Wolpin I, Kadish AH, Wasserstrom JA. Pacing-induced heterogeneities in intracellular Ca²⁺ signaling, cardiac alternans, and ventricular arrhythmias in intact rat heart. *Circ. Res.* 2006; 99: e65-73.
- 74. Bassani JW, Yuan W, Bers DM. Fractional SR Ca release is regulated by trigger Ca²⁺ and SR Ca²⁺ content in cardiac myocytes. *Am. J. Physiol.* 1995; **268**: C1313-9.
- 75. Rovetti R, Cui X, Garfinkel A, Weiss JN, Qu Z. Spark-induced sparks as a mechanism of intracellular calcium alternans in cardiac myocytes. *Circ. Res.* 2010; **106**: 1582-91.

- 76. Nivala M, Qu Z. Calcium alternans in a couplon network model of ventricular myocytes: role of sarcoplasmic reticulum load. *Am. J. Physiol. Heart Circ. Physiol.* 2012; **303**: H341-52.
- 77. Picht E, DeSantiago J, Blatter LA, Bers DM. Cardiac alternans do not rely on diastolic sarcoplasmic reticulum calcium content fluctuations. *Circ. Res.* 2006; **99**: 740-8.
- 78. Comtois P, Nattel S. Atrial repolarization alternans as a path to atrial fibrillation. *J. Cardiovasc. Electrophysiol.* 2012; **23**: 1013-5.
- 79. Narayan SM, Bode F, Karasik PL, Franz MR. Alternans of atrial action potentials during atrial flutter as a precursor to atrial fibrillation. *Circulation* 2002; **106**: 1968-73.
- 80. Rubenstein DS, Lipsius SL. Premature beats elicit a phase reversal of mechanoelectrical alternans in cat ventricular myocytes. A possible mechanism for reentrant arrhythmias. *Circulation* 1995; **91**: 201-14.
- 81. Pastore JM, Girouard SD, Laurita KR, Akar FG, Rosenbaum DS. Mechanism linking T-wave alternans to the genesis of cardiac fibrillation. *Circulation* 1999; **99**: 1385--94
- 82. Qu Z, Garfinkel A, Chen PS, Weiss JN. Mechanisms of discordant alternans and induction of reentry in simulated cardiac tissue. *Circulation* 2000; **102**: 1664-70.
- 83. Cutler MJ, Wan X, Plummer BN, Liu H, Deschenes I, Laurita KR, Hajjar RJ, Rosenbaum DS. Targeted sarcoplasmic reticulum Ca²⁺ ATPase 2a gene delivery to restore electrical stability in the failing heart. *Circulation* 2012; **126**: 2095-104.
- 84. Clusin WT. Mechanisms of calcium transient and action potential alternans in cardiac cells and tissues. *Am. J. Physiol. Heart Circ. Physiol.* 2008; **294**: H1-H10.
- 85. Laurita KR, Rosenbaum DS. Cellular mechanisms of arrhythmogenic cardiac alternans. *Prog. Biophys. Mol. Biol.* 2008; **97**: 332-47.
- 86. Myles RC, Burton FL, Cobbe SM, Smith GL. The link between repolarisation alternans and ventricular arrhythmia: does the cellular phenomenon extend to the clinical problem? *J. Mol. Cell Cardiol.* 2008; **45**: 1-10.
- 87. Altamirano J, Bers DM. Voltage dependence of cardiac excitation-contraction coupling: unitary Ca²⁺ current amplitude and open channel probability. *Circ. Res.* 2007; **101**: 590-7.
- 88. Sheehan KA, Blatter LA. Regulation of junctional and non-junctional sarcoplasmic reticulum calcium release in excitation-contraction coupling in cat atrial myocytes. *J. Physiol.* 2003; **546**: 119-35.
- 89. Fox JJ, McHarg JL, Gilmour RF, Jr. Ionic mechanism of electrical alternans. *Am. J. Physiol. Heart Circ. Physiol.* 2002; **282**: H516-30.
- 90. Shiferaw Y, Watanabe MA, Garfinkel A, Weiss JN, Karma A. Model of intracellular calcium cycling in ventricular myocytes. *Biophys. J.* 2003; **85**: 3666-86.

- 91. Li Y, Diaz ME, Eisner DA, O'Neill S. The effects of membrane potential, SR Ca²⁺ content and RyR responsiveness on systolic Ca²⁺ alternans in rat ventricular myocytes. *J. Physiol.* 2009; **587**: 1283-92.
- 92. Diaz ME, Eisner DA, O'Neill SC. Depressed ryanodine receptor activity increases variability and duration of the systolic Ca²⁺ transient in rat ventricular myocytes. *Circ. Res.* 2002; **91**: 585-93.
- 93. Diaz ME, O'Neill SC, Eisner DA. Sarcoplasmic reticulum calcium content fluctuation is the key to cardiac alternans. *Circ. Res.* 2004; **94**: 650-6.
- 94. Belevych AE, Terentyev D, Viatchenko-Karpinski S, Terentyeva R, Sridhar A, Nishijima Y, Wilson LD, Cardounel AJ, Laurita KR, Carnes CA, Billman GE, Gyorke S. Redox modification of ryanodine receptors underlies calcium alternans in a canine model of sudden cardiac death. *Cardiovasc. Res.* 2009; 84: 387-95.
- 95. Chudin E, Goldhaber J, Garfinkel A, Weiss J, Kogan B. Intracellular Ca²⁺ dynamics and the stability of ventricular tachycardia. *Biophys. J.* 1999; **77**: 2930-41.
- 96. Wan X, Laurita KR, Pruvot EJ, Rosenbaum DS. Molecular correlates of repolarization alternans in cardiac myocytes. *J. Mol. Cell Cardiol.* 2005; **39**: 419-28.
- 97. Eisner DA, Diaz ME, Li Y, O'Neill SC, Trafford AW. Stability and instability of regulation of intracellular calcium. *Exp. Physiol.* 2005; **90**: 3-12.
- 98. Kameyama M, Hirayama Y, Saitoh H, Maruyama M, Atarashi H, Takano T. Possible contribution of the sarcoplasmic reticulum Ca²⁺ pump function to electrical and mechanical alternans. *J. Electrocardiol.* 2003; **36**: 125-35.
- 99. Xie LH, Sato D, Garfinkel A, Qu Z, Weiss JN. Intracellular Ca alternans: coordinated regulation by sarcoplasmic reticulum release, uptake, and leak. *Biophys. J.* 2008; **95**: 3100-10.
- 100. Cutler MJ, Wan X, Laurita KR, Hajjar RJ, Rosenbaum DS. Targeted SERCA2a gene expression identifies molecular mechanism and therapeutic target for arrhythmogenic cardiac alternans. Circ. Arrhythm. Electrophysiol. 2009; 2: 686-94.
- 101. Gwathmey JK, Yerevanian AI, Hajjar RJ. Cardiac gene therapy with SERCA2a: from bench to bedside. *J. Mol. Cell Cardiol.* 2011; **50**: 803-12.
- 102. Lyon AR, Bannister ML, Collins T, Pearce E, Sepehripour AH, Dubb SS, Garcia E, O'Gara P, Liang L, Kohlbrenner E, Hajjar RJ, Peters NS, Poole-Wilson PA, Macleod KT, Harding SE. SERCA2a gene transfer decreases sarcoplasmic reticulum calcium leak and reduces ventricular arrhythmias in a model of chronic heart failure. Circ. Arrhythm. Electrophysiol. 2011; 4: 362-72.
- 103. Brochet DX, Yang D, Di Maio A, Lederer WJ, Franzini-Armstrong C, Cheng H. Ca²⁺ blinks: rapid nanoscopic store calcium signaling. *Proc. Natl.*

- Acad. Sci. USA 2005; 102: 3099-104.
- 104. Cheng H, Lederer MR, Lederer WJ, Cannell MB. Calcium sparks and $[Ca^{2+}]_i$ waves in cardiac myocytes. *Am. J. Physiol.* 1996; **270**: C148-59.
- 105. Ramay HR, Liu OZ, Sobie EA. Recovery of cardiac calcium release is controlled by sarcoplasmic reticulum refilling and ryanodine receptor sensitivity. *Cardiovasc. Res.* 2011; **91**: 598-605.
- 106. Sham JS, Song LS, Chen Y, Deng LH, Stern MD, Lakatta EG, Cheng H. Termination of Ca²⁺ release by a local inactivation of ryanodine receptors in cardiac myocytes. *Proc. Natl. Acad. Sci. USA* 1998; **95**: 15096-101.
- 107. Sobie EA, Song LS, Lederer WJ. Local recovery of Ca²⁺ release in rat ventricular myocytes. *J. Physiol.* 2005; **565**: 441-7.
- 108. Szentesi P, Pignier C, Egger M, Kranias EG, Niggli E. Sarcoplasmic reticulum Ca²⁺ refilling controls recovery from Ca²⁺-induced Ca²⁺ release refractoriness in heart muscle. *Circ. Res.* 2004; **95**: 807-13.
- 109. Sobie EA, Song LS, Lederer WJ. Restitution of Ca²⁺ release and vulnerability to arrhythmias. *J. Cardiovasc. Electrophysiol.* 2006; **17 Suppl 1**: S64-S70.
- 110. Kornyeyev D, Reyes M, Escobar AL. Luminal Ca²⁺ content regulates intracellular Ca²⁺ release in subepicardial myocytes of intact beating mouse hearts: effect of exogenous buffers. *Am. J. Physiol. Heart Circ. Physiol.* 2010; **298**: H2138-53.
- 111. Alvarez-Lacalle E, Cantalapiedra IR, Penaranda A, Cinca J, Hove-Madsen L, Echebarria B. Dependency of calcium alternans on ryanodine receptor refractoriness. *PLoS One* 2013; **8**: e55042. doi: 10.1371/journal.pone.0055042
- 112. Zima AV, Blatter LA. Redox regulation of cardiac calcium channels and transporters. *Cardiovasc. Res.* 2006; **71**: 310-21.
- 113. Aon MA. Mitochondrial dysfunction, alternans, and arrhythmias. *Front. Physiol.* 2013; **4**: 83.

Received 1 July 2013, in revised form 21 October 2013. Accepted 22 October 2013.

© L.A. Blatter 2013.

Author for correspondence:

Lothar A. Blatter
Department of Molecular Biophysics & Physiology
Rush University Medical Center
1750 W. Harrison Street
Chicago, IL 60612
USA

Tel: +1 312 563 3238 Fax: +1 312 942 8711

E-mail: Lothar_Blatter@rush.edu