Post-exercise cold-water immersion activates acute PHF20 and p53 signalling in human skeletal muscle

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Despite its widespread use in post-exercise recovery (Broatch *et al.*, 2014), debate currently exists surrounding the merit of cold-water immersion (CWI) in athletic training regimes. Short-term improvements in recovery from exercise may be thwarted by unfavourable long-term skeletal muscle adaptations (Yamane *et al.*, 2006). The aim of this study was to investigate the underlying molecular mechanisms by which CWI may alter the signalling pathways associated with mitochondrial biogenesis following an acute bout of high-intensity interval exercise.

Nineteen males (mean ± SD; age 24 ± 6 y; \dot{VO}_{2peak} 46.5 ± 8.1 mL•kg⁻¹•min⁻¹) performed an acute highintensity interval training (HIT) bout, comprising 4 × 30s all-out efforts on a cycle ergometer, immediately followed by one of two 15 min recovery conditions: CWI (10.3 ± 0.2°C) or a passive control at ambient room temperature (CON; 23 ± 0.1°C). Muscle biopsies (*vastus lateralis*) were obtained pre-exercise, post-recovery and 3 h post-recovery to determine the acute molecular signalling response following HIT and CWI. Phosphorylation (p-) of p38 MAPK^{Thr180/182} (3.0 ± 0.9 vs 2.4 ± 0.6), AMPK^{Thr172} (2.7 ± 0.8 vs 5.5 ± 1.6) and p53^{Ser15} (1.9 ± 0.4 vs 3.6 ± 1.0) increased immediately post-recovery (P < 0.05) in CON and CWI, respectively. p-p38 MAPK returned to basal levels by 3 hours post-recovery, whereas p-AMPK (2.3 ± 0.5 vs 6.5 ± 2.6) and pp53 (1.6 ± 0.3 vs 4.8 ± 1.5) remained significantly elevated for both conditions (P < 0.05). When compared with CON, CWI resulted in larger increases in p-p53 (ES = 0.92, p = 0.058) and the content of its upstream regulator PHF20 (P < 0.05) immediately post-recovery and 3 h post-recovery (Figure).



Total PHF20 protein and phosphorylation of $p53^{Ser15}$ immediately pre-exercise (Pre), post-recovery (Post), and 3 h post recovery (3h) for CON (open bars) and CWI (closed bars) conditios. *Significant difference from pre-exercise (P < 0.05). **Significant difference from CON. #Large effect (ES > 0.8) from CON. Data are presented as mean \pm S.E.M.

We provide novel data demonstrating that post-exercise CWI alters acute molecular signalling pathways associated with mitochondrial biogenesis. Recently implicated as an important regulator of mitochondrial function (Saleem *et al.*, 2011), p53 activation following CWI may serve as a novel and potent stimulus by which to enhance contraction-induced mitochondrial biogenesis. Our findings are consistent with reports of post-exercise CWI increasing the expression of peroxisome proliferator-activated receptor gamma (PPAR γ) coactivator 1-alpha (PGC-1 α) (Ihsan *et al.*, 2014). The mechanisms by which increases in PHF20 and p53occur following CWI are currently unknown, but may be related to the cellular stress imposed by a hypothermic shock and subsequent rewarming.

Broatch JR, Petersen A, Bishop DJ. (2014) Medicine & Science in Sports & Exercise, In press.

Ihsan M, Watson G, Choo HC, Lewandowski P, Papazzo A, Cameron-Smith D. Abbiss CR. (2014) *Medicine & Science in Sports & Exercise*, **46**: 1900-1907.

Saleem A, Carter HN, Iqbal S. Hood DA. (2011) Exercise and Sport Sciences Reviews, 39: 199-205.

Yamane M, Teruya H, Nakano M, Ogai R, Ohnishi N. Kosaka M. (2006) European Journal of Applied Physiology, 96: 572-580.

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