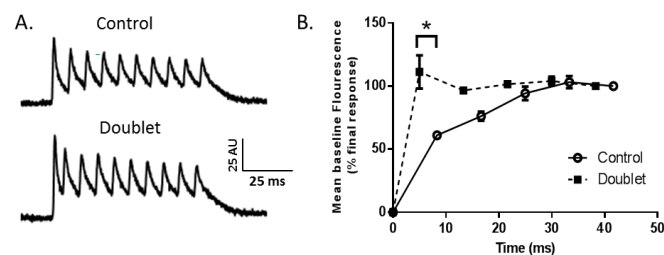


## Effect of doublet stimulation on tetanic $\text{Ca}^{2+}$ responses measured in isolated fast interosseous fibres of the mouse

A.J. Bakker,<sup>1</sup> T.R. Cully<sup>2</sup> and B.S. Launikonis,<sup>2</sup> <sup>1</sup>School of Anatomy, Physiology & Human Biology, University of Western Australia, Perth, WA 6009, Australia and <sup>2</sup>School of Biomedical Sciences, University of Queensland, St Lucia, QLD 4072, Australia.

At the start of tetanic contractions, mammalian skeletal muscle fibres are often excited by high frequency motorneuron double discharges, of 200 Hz or more (Desmedt & Godaux, 1977). Recently, Cheng *et al.*, 2013 reported that the presence of an initial 200 Hz doublet action potential can result in a 100% increase in peak tetanic  $\text{Ca}^{2+}$  compared to control  $\text{Ca}^{2+}$  transients elicited at a stimulation rate of 70Hz alone, there was also a faster rise in force (Cheng *et al.*, 2013). However, Cheng *et al.* tracked  $\text{Ca}^{2+}$  release with only ms temporal resolution, leaving the evolution of the  $\text{Ca}^{2+}$  transient that underlies the rapidly rising force response unknown. In this study, we imaged  $\text{Ca}^{2+}$  release with  $\mu\text{s}$  resolution using a Zeiss 5 Live during doublet action potential stimulation in conjunction with the fast, low affinity  $\text{Ca}^{2+}$  indicator Mag-Fluo-4.

Mice were killed by cervical dislocation, and the interosseous muscles removed and placed in a collagenase II digestion solution (mg/ml) for 30 min. The muscles were then gently triturated to produce single fibres. Fibres were loaded with Mag-Fluo-4 (5 $\mu\text{M}$  for 15 min), and placed in a solution bath containing a HEPES based Krebs's Ringer, and the myosin inhibitor BTS (100  $\mu\text{M}$ ) to prevent fibre movement. Fast fibres were selected based on the shape of elicited tetanic  $\text{Ca}^{2+}$  transients (Calderon, Bolanos & Caputo, 2011). In control measurements, fibres were activated by 10 action potentials at 120 Hz using platinum electrodes. Stimulation during the doublet measurements was similar with the exception that the first 2 action potentials were at 200 Hz.  $\text{Ca}^{2+}$  fluorescence was captured at  $\sim 10$  kHz using a Zeiss 5 Live confocal microscope in linescan mode. Changes in the progression of the  $\text{Ca}^{2+}$  response was quantitated by estimating the lowest  $\text{Ca}^{2+}$  fluorescence found after each  $\text{Ca}^{2+}$  spike, and normalising the value to the fluorescence nadir after the 6<sup>th</sup> response.



A. Effects of control and doublet stimulation on Mag-fluo-4 fluorescence (AU: arbitrary units). B. Changes in the minimum fluorescence value between  $\text{Ca}^{2+}$  transient spikes before (time 0) and after activation under control conditions or with an initial 120 Hz doublet (\* $P < 0.05$ ).

The presence of an initial 200 Hz doublet action potential did not significantly alter the amplitudes of the  $\text{Ca}^{2+}$  spikes during the transient. However, after doublet stimulation, the minimum fluorescence value between  $\text{Ca}^{2+}$  transient spikes rose to a stable, maximal level after the first response that was approximately 1.8 times greater than the basal  $\text{Ca}^{2+}$  fluorescence after the first response under control conditions (control:  $61.06 \pm 2.82\%$  of 6<sup>th</sup> response; doublet:  $111.27 \pm 13.19\%$  of 6<sup>th</sup> response) (Figure). Furthermore, in controls, the minimum fluorescence value between  $\text{Ca}^{2+}$  transient spikes did not reach the maximum normalised value until 16 - 25 ms after initial activation, compared to only 5 ms in fibres exposed to doublet stimulation.

These results indicate that doublet stimulation rapidly increases the minimum fluorescence value between  $\text{Ca}^{2+}$  transient spikes in fast twitch muscle fibres. Doublet activation may lead to more rapid saturation of cytosolic  $\text{Ca}^{2+}$  binding sites and therefore, faster initiation of cross-bridge cycling in fast skeletal muscle fibres.

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