

## Inhibitory effect of phenytoin on cardiac RYR2

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Heart failure affects approximately 1-2% of the Australian population (Sahle *et al.*, 2016). Heart failure is associated with changes in  $\text{Ca}^{2+}$  homeostasis in heart (Bers, 2006) including excessive diastolic leakage of calcium ions ( $\text{Ca}^{2+}$ ) from the sarcoplasmic reticulum (SR) through the ryanodine receptor (RyR2) into the cytoplasm. The  $\text{Ca}^{2+}$  leakage leads to a depletion of the SR and diminution of  $\text{Ca}^{2+}$  release during contraction (systole). In failing heart, the diastolic leakage can be prevented by the RyR2 inhibitor, dantrolene without inhibiting  $\text{Ca}^{2+}$  release during systole or affecting  $\text{Ca}^{2+}$  release in normal healthy hearts (Oo *et al.*, 2015; Maxwell *et al.*, 2012). Unfortunately, dantrolene is hepatotoxic (Muehlschlegel & Sims, 2009) and unsuitable for therapeutic use. Both phenytoin and dantrolene belong to the hydantoin class of compounds. The aim of this study was to investigate properties of phenytoin as an alternative RyR2 inhibitor.

Phenytoin (Dilantin<sup>TM</sup>) is used to treat epilepsy (Gallop, 2010). It is known to modulate  $\text{Na}^+$ ,  $\text{K}^+$  and  $\text{Ca}^{2+}$  channels (Gallop, 2010). It decreases hyperexcitability through blocking the  $\text{Na}^+$  current at serum concentrations between 40 and 80  $\mu\text{mol/l}$  (10 to 20  $\mu\text{g/ml}$ ) with an  $\text{IC}_{50}$  of approximately 58  $\mu\text{mol/l}$  (Lang *et al.*, 1993), without causing sedation or interfering with normal central function (Lang, 1993).

RyR2 channels were isolated from sheep heart and incorporated into artificial lipid bilayers. Single channel RyR2 activity was measured in the presence of cytoplasmic solutions containing 0.1  $\text{mmol/l}$   $\text{Ca}^{2+}$  and 2  $\text{mmol/l}$  ATP (vehicle). The cytoplasmic bath was cycled between 1-minute periods of vehicle and solutions containing different concentrations of phenytoin (10, 20, 50, 100, 200 and 500  $\mu\text{mol/l}$ ). These measurements were also carried out with Calmodulin (CaM, 100  $\text{nmol/l}$ ) since it is known that the dantrolene effect required the presence of CaM (Oo *et al.*, 2015).

The  $\text{IC}_{50}$  values of phenytoin at -40mV were  $19 \pm 3 \mu\text{mol/l}$  and  $10 \pm 2 \mu\text{mol/l}$  (mean  $\pm$  SEM) in the presence and absence of CaM, respectively. At +40mV the corresponding  $\text{IC}_{50}$  values were  $16 \pm 3 \mu\text{mol/l}$  and  $15 \pm 2 \mu\text{mol/l}$ . The Hill coefficients (H) of phenytoin inhibition at -40mV in the presence and absence of CaM were  $1.0 \pm 0.5$  and  $1.5 \pm 1.0$ , respectively. At +40mV these values were  $H = 1.1 \pm 0.6$  and  $1.8 \pm 0.8$  in the presence and absence of CaM, respectively. These values are consistent with a single phenytoin-binding site for inhibition.

Our data provide the first indication that phenytoin could effectively inhibit RyR2 mediated release of  $\text{Ca}^{2+}$  in a manner paralleling that of dantrolene. This is an exciting finding as phenytoin has long been used as a therapeutic agent for epilepsy and hence, unlike chronic use of dantrolene, is human safe. Moreover, we showed that the  $\text{IC}_{50}$  of phenytoin in RyR2 is 10-20  $\mu\text{mol/l}$ , which is  $\sim 1/3$  its  $\text{IC}_{50}$  for the  $\text{Na}^+$  channels. Thus, for RyR2 inhibitory purposes, it is likely that it can be applied at lower concentrations than used therapeutically to inhibit  $\text{Na}^+$  channels.

Bers DM (2006). *Biochemistry Journal* **396**(1): e1-3.

Gallop K. (2010). *Emergency Medicine Australasia* **22**: 108–118.

Lang DG, Wang CM & Cooper BR. (1993). *J Pharmacol Exp Ther* **266**: 829-835.

Maxwell JT, Domeier TL, Blatter LA. (2012). *Am J Physiol Heart Circ Physiol* **302**: H953-963.

Muehlschlegel S & Sims JR. (2009). *Neurocrit Care* **10**: 103-115.

Oo YW, Gomez-Hurtado N, Walweel K, van Helden DF, Imtiaz MS, Knollmann BC & Laver DR. (2015). *Molecular Pharmacology* **88**: 57-63.

Sahle BW, Owen AJ, Mutowo MP, Krum H & Reid CM. (2016). *BMC Cardiovasc Disord* **16**, 32.