



IP₃R activity increases propensity of RyR-mediated Ca²⁺ sparks by elevating dyadic [Ca²⁺]

<u>Joshua Chung</u>¹, Agnė Tilūnaitė^{1,2}, David Ladd^{1,2,3}, Hilary Hunt², Christian Soeller⁴, Edmund J. Crampin^{1,2,3}, Stuart T. Johnston^{2,3}, H. Llewelyn Roderick⁵, Vijay Rajagopal^{1,6}

¹ Department of Biomedical Engineering, The University of Melbourne, Melbourne, VIC 3010, Australia

² School of Mathematics and Statistics, The University of Melbourne, Melbourne, VIC 3010, Australia

³ ARC Centre of Excellence in Convergent Bio-Nano Science and Technology, School of Chemical and Biomedical Engineering, The University of Melbourne, Melbourne, VIC 3010, Australia

⁴ Institute of Physiology, University of Bern, Bern, Switzerland

Abstract: 181P

⁵ Laboratory of Experimental Cardiology, Department of Cardiovascular Sciences, KU Leuven, Belgium

⁶ Baker Department of Cardiometabolic Health, The University of Melbourne, Melbourne, VIC 3010, Australia

The heart's pumping action is governed by the concerted contraction and relaxation of individual cardiomyocytes through the excitation-contraction coupling (ECC) process. During ECC, the cell wide Ca^{2+} release responsible for engaging the cardiomyocyte's contractile machinery is composed of elementary Ca^{2+} release units termed Ca^{2+} sparks, which materialize due to the activation of ryanodine receptors (RyRs) primarily located at 10 - 15 nm wide intracellular microdomains called dyads. While RyRs are the primary Ca^{2+} channels responsible for generating the cell-wide Ca^{2+} transients during ECC, Ca^{2+} release via inositol 1,4,5-trisphosphate (IP₃) receptors (IP₃Rs) are also reported in cardiomyocytes and are demonstrated to elicit ECC-modulating effects. It is proposed that IP₃Rs' ability to modulate ECC is granted by their colocalization with RyRs at functionally relevant sites in the cardiomyocyte, of which includes dyads. Several studies have indeed reported an increase in Ca^{2+} spark frequency under IP₃R stimulation (1–3). However, the mechanism underlying this observation is not fully resolved.

In this study, we aim to uncover the mechanism by which dyad-localized IP₃Rs influence Ca²⁺ sparks and reveal their effect on local Ca²⁺ handling. To this end, we utilized mathematical models of RyRs and IP₃Rs and developed a spatial computational model of the dyad to simulate an environment where clusters of both Ca²⁺ channels are colocalized. Consistent with published experimental data, our biophysics-based simulations predict that this hetero channel crosstalk increases the propensity for RyR-mediated Ca²⁺ spark formation. The stochasticity of IP₃R gating is a key feature to eliciting this outcome. In terms of local Ca²⁺ handling, dyadic IP₃R activity lowers the Ca²⁺ available in the junctional sarcoplasmic reticulum (JSR) for release, thus resulting in Ca²⁺ sparks with lower amplitudes but similar durations. Overall, our results support the hypothesis that IP₃Rs facilitate Ca²⁺ spark formation by raising dyadic Ca²⁺ concentration ([Ca²⁺)), thereby priming RyRs for future activation.

^{1.} Demydenko K, Sipido KR, Roderick HL. Ca2+ release via InsP3Rs enhances RyR recruitment during Ca2+ transients by increasing dyadic [Ca2+] in cardiomyocytes. Journal of Cell Science [Internet]. 2021 Jul 22 [cited 2021 Sep 22];134(14). Available from: https://doi.org/10.1242/jcs.258671

^{2.} Blanch i Salvador J, Egger M. Obstruction of ventricular Ca2+-dependent arrhythmogenicity by inositol 1,4,5-trisphosphate-triggered sarcoplasmic reticulum Ca2+ release. The Journal of Physiology. 2018;596(18):4323–40.

^{3.} Domeier TL, Zima AV, Maxwell JT, Huke S, Mignery GA, Blatter LA. IP3 receptor-dependent Ca2+ release modulates excitationcontraction coupling in rabbit ventricular myocytes. American Journal of Physiology-Heart and Circulatory Physiology. 2008 Feb;294(2):H596–604.